Oxytmatrine suppresses proliferation and facilitates apoptosis of human ovarian cancer cells through upregulating microRNA-29b and downregulating matrix metalloproteinase-2 expression

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Abstract. Oxytmatrine, an alkaloid extracted from medicinal plants of the genus Sophora, has a wide range of pharmacological effects. Previous studies have revealed that oxytmatrine can inhibit proliferation and metastasis of tumor cells through reducing matrix metalloproteinase-2 (MMP-2) mRNA expression. However, the expression of MMP-2 in ovarian cancer is significantly higher than that in normal ovaries. Furthermore, the expression of microRNA-29b (miR-29b) in ovarian carcinoma is significantly lower than that in normal ovaries. Therefore, MMP-2 and miR-29b are tumor suppressor factors involved in ovarian cancer. To evaluate the anti-cancer effects of oxytmatrine the OVCAR-3 ovary cancer cell line was treated with oxytmatrine at the concentrations of 0, 0.5, 1 and 2 mg/ml. Assessment of the proliferation and apoptosis of OVCAR-3 cells showed that oxytmatrine had an inhibitory effect on ovarian cancer cells. Furthermore, oxytmatrine decreased the protein levels of MMP-2 and increased the expression levels of miR-29b in OVCAR-3 cells. Through transfection of miR-29b precursor into OVCAR-3 cells, it was demonstrated that miR-29b regulated MMP-2 expression in OVCAR-3 cells. In addition, anti-miR-29b antibodies were used to verify that the apoptotic effect of oxytmatrine was due to upregulating miR-29b and downregulating MMP-2 expression. These results showed that oxytmatrine suppresses the proliferation and facilitates apoptosis of human ovarian cancer cells through upregulating miR-29b and downregulating MMP-2 expression.

Introduction

Ovarian cancer is a type of malignant tumor which has the highest lethality rate among all gynecological diseases, and possesses high invasion ability as well a high potential to form distant metastases (1). The five-year survival rate of patients is as low as 25-30%. Due to the insidious onset and the asymptomatic characteristics in the early stages, 70-80% of the patients are diagnosed at a late stage, and therefore, the mortality rate ranks first among all gynecological malignancies (2,3). As a part of the hypothalamus-pituitary-ovarian gonadal axis, the ovaries themselves are subject to the regulation of upstream gonadotropins and gonadotropin-releasing hormone, and therefore, regulation of hormone levels is an important mechanism to adjust the incidence of ovarian cancer (4,5).

Matrix metalloproteinase-2 (MMP-2) degrades vascular basement membrane and extracellular matrix, which are major barriers against cancer cell invasion and metastasis; upregulation of MMP-2 in cancer enables tumor cells to break through the barriers and promotes local invasion and distant metastasis of tumor cells (6,7). The mRNA and protein expression levels of MMP-2 in epithelial ovarian cancer are significantly higher than those in normal ovaries, benign ovarian tumors and borderline ovarian tumors (8). MMP-2 overexpression was shown to promote ovarian cancer invasion and metastasis (9).

MicroRNAs (miRs) are short non-coding RNAs, which regulate gene expression in transcription and translation and are involved in a series of important biological processes, including embryonic development in the early stage, cell proliferation, differentiation, apoptosis, fat metabolism and gene expression regulation (10,11). miR-29 has an important role in tumorigenesis, differentiation and tumor development (12). It has been proven that miR-29 expression is downregulated in various human tumor tissues, including lung cancer, hepatocellular carcinoma and prostate cancer. miR-29 has three sub-units: miR-29a, miR-29b and miR-29c (13). miR-29b has been proved to be expressed in breast cancer tissues (14). Expression of miR-29b in epithelial ovarian carcinoma is significantly reduced compared with that in benign ovarian tumor or normal ovarian tissues (15). miR-29b as a tumor suppressor may be involved in epithelial ovarian cancer, as the expression of miR-29b gradually decreases with increasing...
stage; the expression of miR-29b in stage-III-IV epithelial ovarian cancer tissue was shown to be significantly lower than that in stage-I-II ovarian cancer (16).

Oxymatrine (Fig. 1) is an alkaloid extracted from medicinal plants of the genus Sophora, which include bitter beans and broad bean roots, and has a wide range of pharmacological activities, including anti-bacterial, anti-inflammatory, anti-rheumatic, anti-tumor, anti-allergic and immunomodulatory effects (17,18). A large number of studies suggested that oxymatrine can inhibit the proliferation and metastasis of tumor cells, induce apoptosis, cause normal cell differentiation and have anti-tumor activity (17,19-21). Oxymatrine is able to inhibit pancreatic cancer cell invasion and metastasis through significantly reducing MMP-2 mRNA expression (19,22). Furthermore, oxymatrine is able to significantly inhibit liver cancer cell proliferation, induce apoptosis and thus down-regulate the expression of signal transducer and activator of transcription 3 (Stat3) and Stat5 mRNA (23).

The present study investigated the inhibitory role of oxymatrine in ovarian cancer cells and studied the underlying molecular mechanisms. For this, the roles of miR-29b and MMP-2 in the effect of oxymatrine on the proliferation and apoptosis of human ovarian cancer cells were investigated.

Materials and methods

Chemicals and reagents. Oxymatrine (purity, 98%; Sigma-Aldrich, St Louis, MO, USA) was dissolved in physiological saline. Dulbecco's modified Eagle's medium (DMEM) and fetal calf serum (FBS) were obtained from Gibco-BRL (Invitrogen Life Technologies, Carlsbad, CA, USA) and Hyclone (GE Healthcare, Little Chalfont, UK), respectively. MTT was purchased from Sigma-Aldrich.

Cell culture. The human ovarian cancer cell line OVCAR-3 was obtained from the Shanghai Cell Bank of the Chinese Academy of Sciences (Shanghai, China). The cells were cultured in medium containing 10% (v/v) FBS and 100 U penicillin/streptomycin (Invitrogen Life Technologies). Cultured cells were maintained at a temperature of 37˚C in a humidified atmosphere with 5% CO₂. OVCAR-3 cells were detected using a FACSCanto II flow cytometer (Becton Dickinson).

Cell viability assay. The viability of OVCAR-3 cells was determined using the MTT assay. OVCAR-3 cells (1x10⁴ cells/well) were seeded on 96-well plates at a temperature of 37˚C in a humidified atmosphere with 5% CO₂. OVCAR-3 cells were treated with various concentrations of oxymatrine (0, 0.5, 1 and 2 mg/ml). In accordance with the manufacturer's instructions, the cells were washed with PBS and collected in buffer solution (BD Biosciences, Franklin Lakes, NJ, USA). The cells were then incubated with Annexin-V-Fluorescein isocyanate (FITC) and propidium iodide for 30 min on ice. Following incubation, the apoptotic cells were detected using a FACSCanto II flow cytometer (Becton Dickinson).

Flow cytometry. OVCAR-3 cells (1.0-2.0x10⁵ cells/well) were plated in six-well plates and incubated for 24 h. Subsequently, the cells were treated with various concentrations of oxymatrine (0, 0.5, 1 and 2 mg/ml). In accordance with the manufacturer's instructions, the MMP-2 protein levels of OVCAR-3 cells were analyzed by gelatin zymography assays. 20 µl of collected media were added to a fresh centrifuge tube with an equal volume of SDS sample buffer (BD Biosciences). The miscible liquids were subjected to 10% SDS-PAGE (Beyotime Institute of Biotechnology, Jiangsu, China) using an electrophoresis gel impregnated with 0.1% gelatin (Sigma-Aldrich) After electrophoresis, the gel was washed with 2.5% Triton X-100 for 0.5-1 h and incubated in a reaction buffer at 37˚C for 12 h. After incubation, the gel was stained with 0.05% Coomassie brilliant blue R-250 (Amresco, Solon, OH, USA).

Quantitative polymerase chain reaction (qPCR) analysis of miR-29b expression. OVCAR-3 cells were plated on six-well plates (1.0-2.0x10⁵ cells/well) and incubated for 24 h. Then cells were treated with various concentrations of oxymatrine (0, 0.5, 1 and 2 mg/ml). Total RNA was isolated from the cells using TRIzol reagent (Invitrogen Life Technologies), according to the manufacturer's instructions. The RNA was subsequently reverse transcribed into cDNA using the PrimeScript RT Master mix (Takara, Otsu, Japan). In
accordance with the manufacturer's instructions, the miR-29b expression levels were detected using the Bulge-Loop miRNA qRT-PCR kit (Guangzhou Ribobio, Guangzhou, China). Cycling conditions were as follows: 95˚C for 10 min followed by 35 cycles of 95˚C for 15 sec, 56˚C for 30 sec and 72˚C for 30 sec, and a final elongation at 72˚C for 7 min, on an ABI 7500 system (Takara). For miR-29b, the primers were as follows: 5'-ACGCAAATTCGTGAAGCGTT-3' and 5'-UAGCACCAUUUGAAAUCAGUGUU-3'. The primers for β-actin were as follows: 5'-GTGGACATCCGTAAAGACC-3' and 5'-GGAGCAGGGCAGTAA-3'.

Transfection of miR-29b and anti-miR-29b. The pcDNA-miR-29b precursor and pcDNA-anti-miR-29b were synthesized by Ribobio Biological Tecnology Co. (Shanghai, China). A total of 100 nmol/l miR-29b or anti-miR-29b was transfected into the cells with Lipofectamine 2000 (Invitrogen Life Technologies). Twenty-four hours after transfection, cells were treated with various concentrations of oxymatrine (0, 0.5, 1 and 2 mg/ml).

Statistical analysis. Statistical analysis was performed with SPSS 17.0 software (SPSS, Inc., Chicago, IL, USA). Differences were assessed using analysis of variance or Student's t-test. Values are expressed as the mean ± standard deviation. P<0.05 was considered to indicate a statistically significant difference between values.

Results

Oxymatrine inhibits OVCAR-3 cell growth and leads to the activation of caspase-3. In the present study, the proliferation of OVCAR-3 cells was measured by using the MTT assay. After treatment with oxymatrine (0, 0.5, 1 and 2 mg/ml) (24,25), the proliferation of OVCAR-3 cells was reduced (Fig. 2A). Treatment with oxymatrine at the concentrations of 1 and 2 mg/ml significantly inhibited the proliferation at 48 h and 72 h (P<0.01) (Fig. 2A). Furthermore, treatment with oxymatrine at the concentrations of 1 and 2 mg/ml at 48 h, increased the activity of caspase-3 (Fig. 2B). Treatment with oxymatrine at the concentrations of 1 and 2 mg/ml significantly increased caspase activity (P<0.01) (Fig. 2B).

Oxymatrine induces apoptosis in ovarian cancer cells. After treatment with oxymatrine (0, 0.5, 1 and 2 mg/ml), the apoptotic effect of OVCAR-3 cells was measured by staining...
with Annexin-V-FITC/propidium iodide for 48 h followed by flow-cytometric analysis. Treatment with oxymatrine at the concentrations of 1 and 2 mg/ml significantly increased the apoptotic effect (P<0.01) (Fig. 3).

**Oxymatrine decreases the expression of MMP-2.** After treatment with oxymatrine (0, 0.5, 1 and 2 mg/ml) for 48 h, the protein levels of MMP-2 in OVCAR-3 cells were assessed using gelatin zymography assays (Fig. 4A). Treatment with oxymatrine at the concentrations of 1 and 2 mg/ml significantly decreased the protein levels of MMP-2 (P<0.01) (Fig. 4B).

**Oxymatrine stimulates miR-29b expression.** After treatment with oxymatrine (0, 0.5, 1 and 2 mg/ml), the miR-29b expression levels of OVCAR-3 cells were assessed using PCR analysis at 48 h. The miR-29b expression levels of OVCAR-3 cells were increased (Fig. 5). Treatment with oxymatrine at the concentrations of 1 and 2 mg/ml significantly increased the miR-29b expression levels (P<0.01) (Fig. 5).

**Overexpression of miR-29b decreases MMP-2 expression.** To determine whether miR-29b regulates MMP-2 expression in OVCAR-3 cells, the protein levels of MMP-2 in OVCAR-3 cells were assessed following transfection of the cells with miR-29b precursor. The effect of miR-29b precursor on the miR-29b expression was determined by qPCR analysis. The results indicated that transfection with miR-29b precursor significantly elevated the expression of miR-29b in OVCAR-3 cells (P<0.01) (Fig. 6A) and obviously decreased MMP-2 expression (Fig. 6B).

**The anti-proliferative effect of oxymatrine is mediated via miR-29b, which negatively regulates MMP-2 expression.** The anti-miR-29b vector was employed to investigate the role of miR-29b in the effect of oxymatrine on the growth inhibition of OVCAR-3 cells. The results indicated that anti-miR-29b efficiently penetrated into OVCAR-3 cells and significantly reduced the expression of miR-29b (Fig. 7A). Furthermore, anti-miR-29b significantly reduced the effect of oxymatrine (1 mg/ml) on the apoptosis (Fig. 7B) and proliferation (Fig. 7C) of OVCAR-3 cells at 48 h and neutralized the inhibitory effect of oxymatrine (1 mg/ml) through downregulating MMP-2 activity (Fig. 7D) in OVCAR-3 cells.

**Discussion**

To the best of our knowledge, the present study was the first to investigate the inhibitory effects of oxymatrine on ovarian cancer cells. Oxymatrine significantly reduced the proliferation and induced apoptosis of OVCAR-3 cells. Furthermore, oxymatrine significantly decreased the MMP-2 proteins level and increased the miR-29b expression levels in ovarian cancer cells. The results suggested that the apoptotic effect of oxymatrine was due to upregulating miRNA-29b and downregulating MMP-2 expression, confirmed by reversal of this effect following transfection with anti-miR-29b.

Ovarian cancer is a malignant disease with multiple gene mutations and stages; the specific cancer-associated gene mutations are important factors regarding the occurrence, development and prognosis of ovarian cancer (17). Oxymatrine is an alkaloid with numerous pharmacological activities. Oxymatrine inhibits the proliferation of tumor cells by inhibiting their DNA synthesis, affecting their cell cycle (26). MMP-2 is capable of degrading collagen, gelatin, laminin, fibronectin, elastin, proteoglycans and other important components of the extracellular matrix and promoting the invasion, metastasis and angiogenesis of tumor cells through degrading the basement membrane and remodeling influential tissues of the extracellular matrix (7,27).

In the present study, the proliferation of OVCAR-3 cells was significantly inhibited after treatment with oxymatrine (1 and 2 mg/ml) at 48 and 72 h. It was therefore suggested that oxymatrine may be suitable for the treatment of ovarian cancer.
Oxymatrine induces apoptosis of tumor cells by affecting the expression of genes associated with tumor cells and inhibiting the activity of associated enzymes (28). High-level expression of MMP-2 has been found in numerous types of tumor tissue, and is associated with the tumor's invasiveness. Oxymatrine was shown to significantly inhibit the invasion of MDA-MB-231 cells by inhibiting the activation of MMP-2/MMP-9 (22).

Oxymatrine can dose-dependently inhibit the proliferation of cancer cells. As for different types of tumor cell, the difference in the effective concentration is relatively large, with the effective concentration ranging from 0.2 to 16 mmol/l (29). In the present study, the effects of oxymatrine on the apoptosis and caspase-3 activity in ovarian cancer cells were detected using the flow-cytometric analysis and a caspase-3 colorimetric assay kit, respectively. The results showed that oxymatrine (1 and 2 mg/ml) significantly increased the apoptosis of OVCAR-3 cells at 48 h. Furthermore, oxymatrine significantly decreased the MMP-2 protein levels in ovarian cancer cells, as indicated by gelatin zymography assays.

miRNAs are non-coding, single-stranded RNAs with a low molecular weight, which commonly occur in eukaryotic cells (10). Cell proliferation, differentiation and apoptosis are regulated by the degradation of target mRNAs or inhibition of translation, and participate in processes involved in ontogeny, body metabolism and tumor development (12). Members of the miR-29 family were shown to promote the development of breast and colon cancer by inducing apoptosis (30). miR-29b may also increase the sensitivity of cells to apoptosis by inhibiting the expression of myeloid cell leukemia 1 in cholangiocarcinoma cells and promote tumorigenesis, indicating that miR-29 is closely associated with tumor development (30). The expression of miR-29b in epithelial ovarian cancer patients with lymph node metastasis was found to be significantly lower than that in patients without lymph node metastasis, which indicates that the low expression of miR-29b and the metastasis of ovarian cancer are closely associated (15,31). The present study demonstrated that oxymatrine increased the expression of miR-29b in OVCAR-3 cells using qPCR analysis. Furthermore, miR-29b was found to regulate MMP-2 expression of OVCAR-3 cells through transfection of miR-29b precursor into OVCAR-3 cells. In addition, anti-miR-29b was used to verify that the apoptotic effect of oxymatrine was due...
to upregulating miRNA-29b and downregulating MMP-2 expression.

In the present study, oxymatrine treatment (1 and 2 mg/ml for 48 and 72 h) was shown to significantly inhibit the proliferation of OVCAR-3 cells. Furthermore, oxymatrine significantly increased the apoptosis and the activity of caspase-3 in OVCAR-3 cells. In addition, oxymatrine (1 mg/ml) decreased the protein levels of MMP-2 and increased the expression of miR-29b in OVCAR-3 cells at 48 h. Through transfection of miR-29b precursor into OVCAR-3 cells and treatment with oxymatrine (1 mg/ml) for 48 h, it was indicated that miR-29b regulates MMP-2 expression in OVCAR-3 cells. Furthermore, anti-miR-29b was used to verify that the apoptotic effect of oxymatrine was mediated through the upregulation of miR-29b and downregulation of MMP-2 expression after treatment with oxymatrine (1 mg/ml) for 48 h. These results showed that oxymatrine suppresses the proliferation and facilitates apoptosis of human ovarian cancer cells through upregulating miR-29b and downregulating MMP-2 expression.

References


